



# Analysis of mesophyll conductance in five understory herbaceous species

Rosangela Catoni<sup>1</sup> · Francesco Bracco<sup>1</sup> · Mirko U. Granata<sup>1</sup>

Received: 27 February 2019 / Revised: 2 August 2019 / Accepted: 10 December 2019 / Published online: 17 January 2020  
© Prof. H.S. Srivastava Foundation for Science and Society 2020

**Abstract** Mesophyll conductance ( $g_m$ ) has received over time much less attention than stomatal conductance ( $g_s$ ), although it affects leaf photosynthesis to about the same extent as stomatal conductance does. The objective of this study was to analyze the  $g_m$  trend in five understory herbaceous species growing in a close-canopy forest in the north-west of Italy. In particular, three of analyzed species were monocots: *Carex brizoides* Lam., *Carex pilosa* Scop., and *Oplismenus undulatifolius* P. Beauv and the others dicots species: *Circaea lutetiana* L., and *Pulmonaria officinalis* Ced. The results showed, on one hand, the absence of correlation between  $g_m$  and the considered environmental variables in the forest understory (i.e. air temperature, photosynthetic photon flux density and carbon dioxide concentration). Moreover, we carried out a principal component analysis considering all the analyzed morphological and physiological variables for the five species. The following correlation between the first component, related to the leaf mass per unit of leaf area and the leaf tissue density, and  $g_m$  seem to suggest a key role of the leaf structural features in determining  $g_m$  variations across the five species.

**Keywords** Mesophyll conductance · Herbaceous species · Understory species · Functional group

## Introduction

Net photosynthetic rate is affected by several parameters, and among all, stomatal conductance is certainly the best characterized receiving the greatest attention over time (Kumar et al. 1999; Park and Furukawa 1999; Massacci et al. 2008). However, another important factor having a key role as photosynthetic parameter, but just more recently recognized, is the mesophyll conductance (Nascimento and Marengo 2013). Mesophyll conductance ( $g_m$ ) is the ease with which carbon dioxide ( $\text{CO}_2$ ) diffuses from intercellular airspace within a leaf to the sites of carboxylation within chloroplasts, and it constrains photosynthesis to about the same extent as stomatal conductance does (von Caemmerer and Evans 1991; Flexas et al. 2012).

Based on the currently knowledges (review by Flexas et al. 2008) the observed variations of  $g_m$  among different plant groups (i.e. evergreen gymnosperm, evergreen angiosperms, semi-deciduous angiosperms, deciduous angiosperms, herbaceous annual, perennial herbs, CAM plants) are mainly associated to the leaf forms and plant functional group rather than reflecting evolutionary trends. Taking as example the herbaceous group, in this case, the greater differentiation is found between the functional groups of annuals/biannuals versus perennials herbs, while no differences were found along the evolutionary differentiation of dicots versus monocots. Anyway, further the general trend based on the average data per groups, studies show significant variability within a single group, genus and species. In fact, in annual and biannual herbaceous species, the  $g_m$  range can go from  $0.08 \text{ mol CO}_2 \text{ m}^{-2} \text{ s}^{-1} \text{ bar}^{-1}$  to  $g_m$  values over  $1 \text{ mol CO}_2 \text{ m}^{-2} \text{ s}^{-1} \text{ bar}^{-1}$ . This last aspect, about the greater variability within group, genus and species, suggest that  $g_m$  is a parameter that can adapt very quickly and is therefore involved in the

✉ Rosangela Catoni  
rosangela.catoni@unipv.it

<sup>1</sup> Department of Earth and Environmental Sciences, University of Pavia, Via S. Epifanio 14, 27100 Pavia, Italy

differences in photosynthetic efficiency found between species. Besides this large variability among species,  $g_m$  shows a long-term response to different environmental factors (Flexas et al. 2008). Concerning the abiotic factors, as underlined by previous works,  $g_m$  is affected by water stress, low nitrogen availability and salinity (Jones 1973; Evans and Terashima 1988; Bonggi and Loreto 1989). Moreover,  $g_m$  responds to change in light conditions. In fact, shade leaves show a lower  $g_m$  values compared to sun leaves (Hanba et al. 2002; Piel et al. 2002; Laisk et al. 2005; Warren et al. 2007; Catoni et al. 2015a), whilst a complex response in the acclimation to different growth temperature has been suggested (Yamori et al. 2006; Diaz-Espejo et al. 2007). Moreover, referring to internal factors, such as leaf development and ageing, they seem to strongly affect  $g_m$  (Flexas et al. 2008). On one hand, during leaf development (i.e. from unfolding to maturation)  $g_m$  increases in parallel with leaf photosynthesis (Miyazawa and Terashima 2001), while on the other hand, leaf ageing results in a  $g_m$  decreases (Bernacchi et al. 2005; Grassi and Magnani 2005; Catoni et al. 2015a). Finally, a relationship between  $g_m$  and leaf structural traits such as leaf mass per area (LMA) has been reported (De Lucia et al. 2003; Warren et al. 2003).

Based on these observations,  $g_m$  seems to have a long-term response to several environmental factors, and in turn, these changes are important in affecting photosynthesis response to environmental conditions. Nevertheless, others studies reported a stable  $g_m$  in response to changes in environmental variables such as CO<sub>2</sub> concentration, irradiance and temperature (von Caemmerer and Evans 1991; Tazoe et al. 2009; Nascimento and Marengo 2013).

The objective of this study was to analyze, on one hand the  $g_m$  long-term response to the variations of environmental conditions in five herbaceous understory species; and on the other hand, how  $g_m$  varies among these species belonging to different evolutionary groups (dicots and monocots) and functional groups (annual and perennial).

## Materials and methods

### Plant materials and study area

The study was carried out in the period June–October 2018 on five understory herbaceous species. In particular, three monocots: *Carex brizoides* Lam., *Carex pilosa* Scop., and *Oplismenus undulatifolius* P. Beauv and two dicots species: *Circaea lutetiana* L., and *Pulmonaria officinalis* Ced. were considered. All these represent the most important and representative species constituting the herbaceous layer of a typical alluvial lowland forest inside a Strict Natural Reserve “Bosco Siro Negri” (45°12′39″N; 09°03′26″E,

74 m a. s. l.) in Italy. During the study period all the considered species were well representative and without any sign of senescence. Specifically, in the herbaceous layer of the considered forest *C. lutetiana* and *O. undulatifolius* are generally found from the middle of May to beginning of November (i.e. annual species), *C. pilosa* and *P. officinalis* are present throughout the year (i.e. perennial species), while *C. brizoides* seems to have a borderline behavior between annual and perennial herbaceous species, showing up later during the year compared to the two perennial species. For each species five plots (1 m<sup>2</sup> each) were randomly select and inside each plot three representative individual were identify. The structure of the forest is that of a typical closed canopy forest, with a forest tree density of 237 ± 100 stems ha<sup>-1</sup>, a total basal area of 74.5 ± 24.6 m<sup>2</sup> ha<sup>-1</sup> and a Leaf Area Index (LAI) of 4.5 ± 0.3 (Catoni et al. 2015a, b). Moreover, the high tree density determines a high light extinction at soil level, with a value of the relative intercept irradiance by the canopy at soil level inside the forest of 0.77 ± 0.12% and a value of the ratio between irradiance in the red and far red wavelengths (R/FR) of 0.5 ± 0.1 (data from Granata et al. 2016).

The soil is characterized by a pH of 5.46 ± 0.11, a nitrogen concentration (N) of 1.61 ± 0.12 mg g<sup>-1</sup>, a carbon to nitrogen ratio (C/N) of 17 ± 1 and a soil organic matter (SOM) concentration of 44.47 ± 0.04 mg g<sup>-1</sup> (Catoni et al. 2015a).

The climate of the area is characterized by a total annual rainfall of 627 mm most of it falling in autumn and winter. The mean minimum air temperature (T<sub>min</sub>) of the coldest month (January) is 0.1 ± 1.5 °C, the mean maximum air temperature (T<sub>max</sub>) of the hottest month (July) 30.2 ± 1.2 °C and the mean annual temperature (T<sub>m</sub>) 13.8 ± 8.2 °C (Lombardia Regional Agency for Environmental Protection, Meteorological Station of Pavia, Ponte Ticino SS35, data for the period 2002–2017). Floods occurred sporadically every 5–10 years during the last 40 years, with water levels up to 1.50 m height in the forest during exceptional events (Castagneri et al. 2013; Motta et al. 2009). On average, groundwater level is around – 4.50 m in winter reaching – 3.50 m in summer due to irrigation in the surrounding area.

During the study period total rainfall was 256 mm, T<sub>m</sub> 22.2 ± 3.9 °C and T<sub>max</sub> (July and August) 30.7 ± 0.1 °C (Lombardia Regional Agency for Environmental Protection, Meteorological Station of Pavia, Ponte Ticino SS35, data for the period June–October 2018).

### Leaf morphology and ambient conditions

Fully expanded leaves (for each species n = 2 leaves per 3 plant per plot) were collected in June. Leaf samples were

sealed in plastic bags and transported immediately to the laboratory. Measurements included leaf surface area (LA, cm<sup>2</sup>), obtained by the image analysis system (Delta-T Devices, UK) and leaf dry mass (DM, mg), determined by drying leaves at 80 °C to constant mass.

Leaf mass per unit of leaf area (LMA, mg cm<sup>-2</sup>) was calculated by the ratio of DM and LA and leaf tissue density (LTD, mg cm<sup>-3</sup>) was calculated by the ratio of LMA and leaf thickness (LT, μm). This last was determined using digital calipers (precision of 0.01 mm) on the middle portion of the leaf blade and at two different points per leaf avoiding major veins.

During the study period, photosynthetic photon flux density (PPFD), air relative humidity (RH), air temperature (T<sub>air</sub>) and atmospheric CO<sub>2</sub> concentration (CO<sub>2</sub>, ppm) in the forest understory were recorded using specific sensors connected to data loggers.

### Gas exchange

Gas exchange measurements were carried out in the period June–October 2018. Leaves (n = 5 leaves per each species) were retained in their natural position during measurements. Net photosynthetic rate [ $P_N$ , μmol(CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>], stomatal conductance [ $g_s$ , mol(H<sub>2</sub>O) m<sup>-2</sup> s<sup>-1</sup>], leaf transpiration [ $E$ , mmol(H<sub>2</sub>O) m<sup>-2</sup> s<sup>-1</sup>], photosynthetic photon flux density [PPFD, μmol(photons) m<sup>-2</sup> s<sup>-1</sup>] leaf temperature (T<sub>l</sub>, °C), and substomatal CO<sub>2</sub> concentration (C<sub>i</sub>, ppm) were measured by an infrared gas analyzer (LC-Pro +, ADC, UK) equipped with a leaf chamber (PLC, Parkinson Leaf Chamber, UK). On each sampling occasion, leaf respiration [ $R_D$ , μmol(CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>] was measured after  $P_N$  measurements (on the same leaves) as CO<sub>2</sub> efflux by darkening the leaf chamber with a black paper, according to Cai et al. (2005) for 30 min prior to each measurement, to avoid the release of CO<sub>2</sub> transient post-irradiation bursts (Atkin et al. 1998).

### Chl fluorescence and mesophyll conductance

Chl fluorescence measurements were carried out by a portable modulated fluorometer (OS5p, Opti-Sciences, USA). Maximum PSII photochemical efficiency (F<sub>v</sub>/F<sub>m</sub>) was estimated by darkening leaves (n = 10 leaves per each species) for 20 min, then a saturating pulse was applied to measure initial (F<sub>0</sub>) and maximum (F<sub>m</sub>) fluorescence. F<sub>v</sub>/F<sub>m</sub> was estimated as:

$$(F_m - F_0)/F_m$$

Additional gas-exchange and Chl fluorescence measurements were made on leaves (n = 10 leaves per each species) comparable to those previously used in order to

estimate mesophyll conductance [ $g_m$ , mol(CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup> bar<sup>-1</sup>].

The  $g_m$  was calculated according to Harley et al. (1992) by a single point method which combines gas-exchange and Chl a fluorescence measurements, as:

$$g_m = P_N / \{C_i - [\Gamma^* \times [ETR + 8 \times (P_N + R_D)]] / [ETR - 4 \times (P_N + R_D)]\}$$

where  $\Gamma^*$  was the CO<sub>2</sub> compensation point under non respiratory conditions. The temperature dependency for  $\Gamma^*$  was calculated according to Bernacchi et al. (2002).

$P_N$ ,  $C_i$ , and  $R_D$  were obtained from gas-exchange measurements as described in the above section. ETR was the electron transport rate calculated from Chl fluorescence measurements, according to Krall and Edwards (1992) as:

$$ETR = \Phi_{PSII} \times PPFD \times 0.5 \times 0.84$$

where  $\Phi_{PSII}$  was the actual PSII photochemical efficiency of light-adapted leaves calculated according to Genty et al. (1989) as:

$$(F_m' - F_s)/F_m'$$

F<sub>m</sub>' was the maximum fluorescence obtained with a light-saturating pulse (~ 8000 μmol m<sup>-2</sup> s<sup>-1</sup> PPFD) and F<sub>s</sub> was the steady-state fluorescence of illuminated leaves (1500 μmol m<sup>-2</sup> s<sup>-1</sup> PPFD). Gas-exchange and Chl fluorescence measurements were taken concurrently by fitting the portable infrared CO<sub>2</sub> gas analyzer with a fluorometer adapter chamber (F.LCI-FL, ADC, UK).

### Data analysis

The obtained data [expressed as mean ± standard error (± SE)] were analyzed by the analysis of variance (ANOVA). As the same leaves were measure during the course of the study period, data were analyze using a repeated measures analysis of variance. Tukey's post hoc test (p < 0.05) was use to assess the significant differences among the species and between months. Linear regression analysis was carried out between  $g_m$  and the considered environmental variables (i.e. CO<sub>2</sub> concentration, PPFD and T<sub>air</sub>). A principal component analysis (PCA) was carried out in order to summarize the considered morphological (LMA, LTD and LT) and physiological ( $P_N$ ,  $R_D$ ,  $g_m$ ,  $g_s$ , F<sub>v</sub>/F<sub>m</sub>,  $\Phi_{PSII}$ , ETR) leaf traits into major components which explained their variation between the five considered species. Then, a linear regression analysis between the axis explaining the largest proportion of the variance (i.e. PC1) and  $g_m$  was carried out.

## Results

### Leaf morphology and ambient conditions

At morphological level significant differences were found among the considered species (Table 1).

In particular LMA ranged from  $1.55 \pm 0.15 \text{ mg cm}^{-2}$  (*O. undulatifolius*) to  $5.85 \pm 0.48 \text{ mg cm}^{-2}$  (*C. brizoides*) while LTD ranged between  $319.7 \pm 13.8 \text{ mg cm}^{-3}$  (*P. officinalis*) to  $1024.1 \pm 65.1 \text{ mg cm}^{-3}$  (*C. brizoides*). *P. officinalis* showed the highest LT ( $110.0 \pm 8.9 \mu\text{m}$ ) and *O. undulatifolius* and *C. brizoides* the lowest ( $50.65 \pm 0.01 \mu\text{m}$ ). During the study period in the forest understory, mean monthly PPFD ranged between  $51 \pm 5 \mu\text{mol m}^{-2} \text{ s}^{-1}$  in September to  $122 \pm 48 \mu\text{mol m}^{-2} \text{ s}^{-1}$  in June and monthly understory  $T_{\text{air}}$  varied between  $20.9 \pm 0.5 \text{ }^\circ\text{C}$  in October to  $28.4 \pm 0.5 \text{ }^\circ\text{C}$  in July. The average value of  $\text{CO}_2$  concentration during the study period was  $449 \pm 29 \text{ ppm}$ .

### Gas exchange

During the study period the highest  $P_N$  was measured in June for all the considered species, with *C. pilosa* having the highest value [ $4.33 \pm 0.3 \mu\text{mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1}$ ] and *O. undulatifolius* the lowest [ $1.31 \pm 0.06 \mu\text{mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1}$ ] (Fig. 1a). A steady trends of  $R_D$  has been observed (Fig. 1b) with higher rates in July [ $0.56 \pm 0.05 \mu\text{mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1}$ , mean value of the species] decreasing by 11% (mean value) in September–October. Stomatal conductance showed the same  $P_N$  trend, with the highest values in June for all the species [ $0.171 \pm 0.063 \text{ mol}(\text{H}_2\text{O}) \text{ m}^{-2} \text{ s}^{-1}$ , mean value] (Fig. 2).

### Chl fluorescence and mesophyll conductance

Values of  $\Phi_{\text{PSII}}$  and  $F_v/F_M$  are show in Fig. 3a and b. Quite constant trends were monitored during the study period for both the parameters and in all the considered species, with *C. pilosa* having the lowest  $\Phi_{\text{PSII}}$  during the study period

( $0.691 \pm 0.023$ , mean value of the considered months) and *P. officinalis* the highest ( $0.742 \pm 0.003$ , mean value). This last showed also the highest  $F_v/F_M$  ( $0.793 \pm 0.003$ , mean value of the considered months) and *C. lutetiana* the lowest ( $0.765 \pm 0.002$ , mean value).

During the study period (Fig. 4) the highest  $g_m$  value was monitored in July [ $0.015 \pm 0.011 \text{ mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1} \text{ bar}^{-1}$ , mean value], with *P. officinalis* having the highest value. The results of the linear regression showed that none of the considered environmental variables seems to affect  $g_m$  variations during the study period (Table 2).

### Principal component analysis

The PCA highlighted that the first two components accounted for 99.9% of the total variance, with the first component explained the 99.4% of the total variance and it was positively related to LMA ( $r = 0.805$ ) and LTD ( $r = 0.999$ ). The second component explained 0.62% of the total variance was only related to LT ( $r = 0.891$ ). Three different groups can be observed (Fig. 5): one group was represented by *C. pilosa* and *P. officinalis*, and the second group by *O. undulatifolius* and *C. lutetiana*, while *C. brizoides* was placed far from the other two groups. Along the first component *C. brizoides* showed the highest value and the *P. officinalis*, *C. lutetiana* and *O. undulatifolius* the lowest one. There was a significant linear regression between PC1 and  $g_m$  ( $R^2 = 0.24$ ,  $p < 0.01$ ) (Fig. 6).

## Discussion

On the whole, during the analyzed months, in which all the species are well evident and representative and without any sign of senescence, the lowest  $g_m$  rates are observed in June [ $0.012 \pm 0.007 \text{ mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1} \text{ bar}^{-1}$ , mean value of the considered species] and the highest ones in July [ $0.015 \pm 0.011 \text{ mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1} \text{ bar}^{-1}$ , mean value]. Overall, the average  $g_m$  value for the five species over the entire study period [ $0.014 \pm 0.008 \text{ mol}(\text{CO}_2) \text{ m}^{-2} \text{ s}^{-1}$

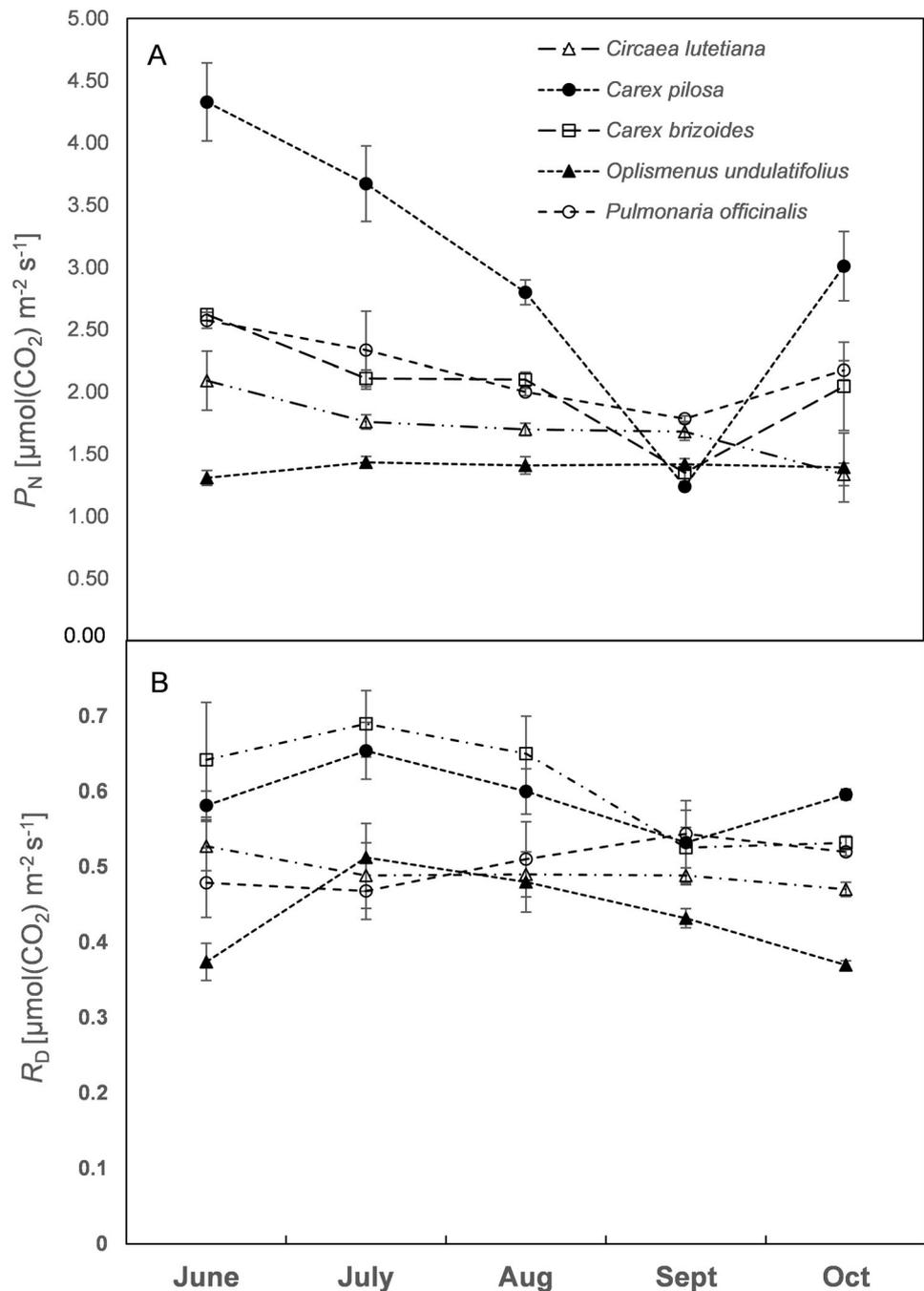
**Table 1** Morphological and anatomical leaf traits of the considered species

	LMA ( $\text{mg cm}^{-2}$ )	LTD ( $\text{mg cm}^{-3}$ )	L ( $\mu\text{m}$ )
<i>Carex brizoides</i>	$5.85 \pm 0.48\text{a}$	$1024.1 \pm 65.1\text{a}$	$50.6 \pm 3.0\text{a}$
<i>Carex pilosa</i>	$4.84 \pm 0.38\text{a}$	$406.3 \pm 16.0\text{b}$	$94.4 \pm 4.4\text{b}$
<i>Circaea lutetiana</i>	$2.01 \pm 0.06\text{b}$	$327.0 \pm 29.5\text{b}$	$63.3 \pm 3.9\text{a}$
<i>Oplismenus undulatifolius</i>	$1.55 \pm 0.15\text{b}$	$324.1 \pm 16.9\text{b}$	$50.7 \pm 4.8\text{a}$
<i>Pulmonaria officinalis</i>	$3.18 \pm 0.06\text{c}$	$319.7 \pm 13.8\text{b}$	$110.0 \pm 8.9\text{b}$

LMA leaf mass per unit of leaf area, LTD leaf tissue density, L leaf thickness

Mean values ( $\pm$  SE) are shown ( $n = 35$  leaves). Different letters indicate significant differences among the species (Tukey-test,  $p < 0.05$ )

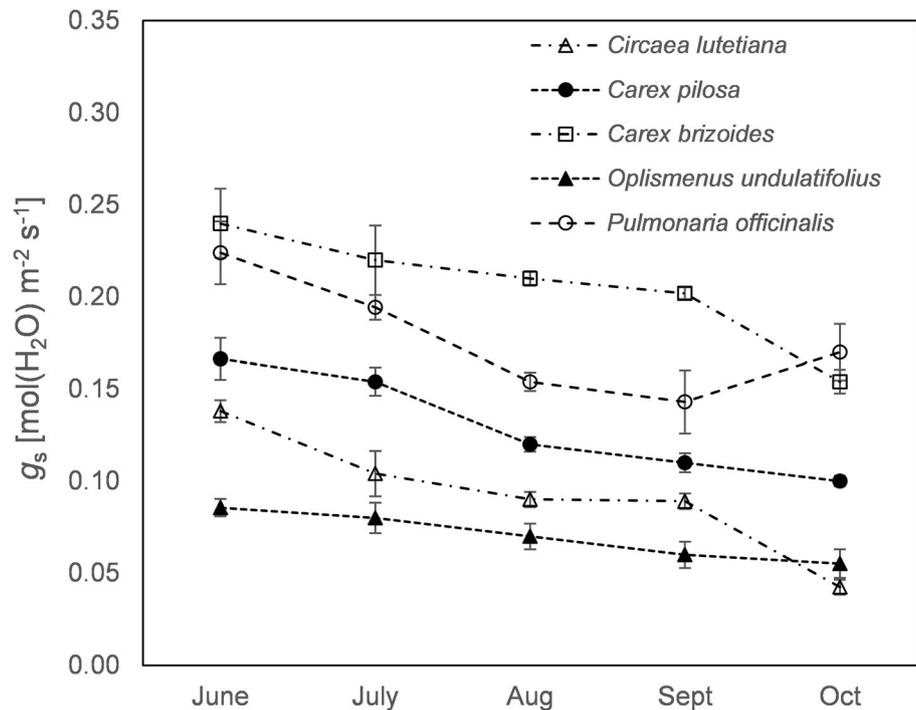
**Fig. 1** Trend of **a** net photosynthetic rates ( $P_N$ ) and **b** dark respiration rates ( $R_D$ ) during the study period in the five considered species. Mean values ( $\pm$  SE) are shown ( $n = 5$ )



$\text{bar}^{-1}$ ] results lower compared to an expected value for herbaceous species (see Flexas et al. 2008). Nevertheless, taking into account the typical shade environment of the forest understory in which they grow, this value may not be surprising considering that normally shade leaves have lower  $g_m$  values than leaves in high-light environments. In fact, understory species might behave similarly to shade-leaves, which appear to have stronger limitations from  $g_m$  (Tosens et al. 2012; Cano et al. 2013). As suggested by Laisk et al. (2005) the higher mesophyll diffusion

resistance (i.e. lower  $g_m$ ) can reflect the smaller air-exposed mesophyll area in the thinner shade leaves. Moreover, according to the results of Taylor and Pearcy (1976) for understory species, the observed  $g_m$  rates are substantially lower compared to  $g_s$ . Another typical behavior observed in the understory environment is the photosynthetic response that generally parallels the change in the light environment induced by overstory canopy closure. In fact, the highest  $P_N$  rates are found, in all these species, in June prior to the complete canopy closure, with a further

**Fig. 2** Trend of stomatal conductance ( $g_s$ ) during the study period in the five considered species. Mean values ( $\pm$  SE) are shown ( $n = 5$ )



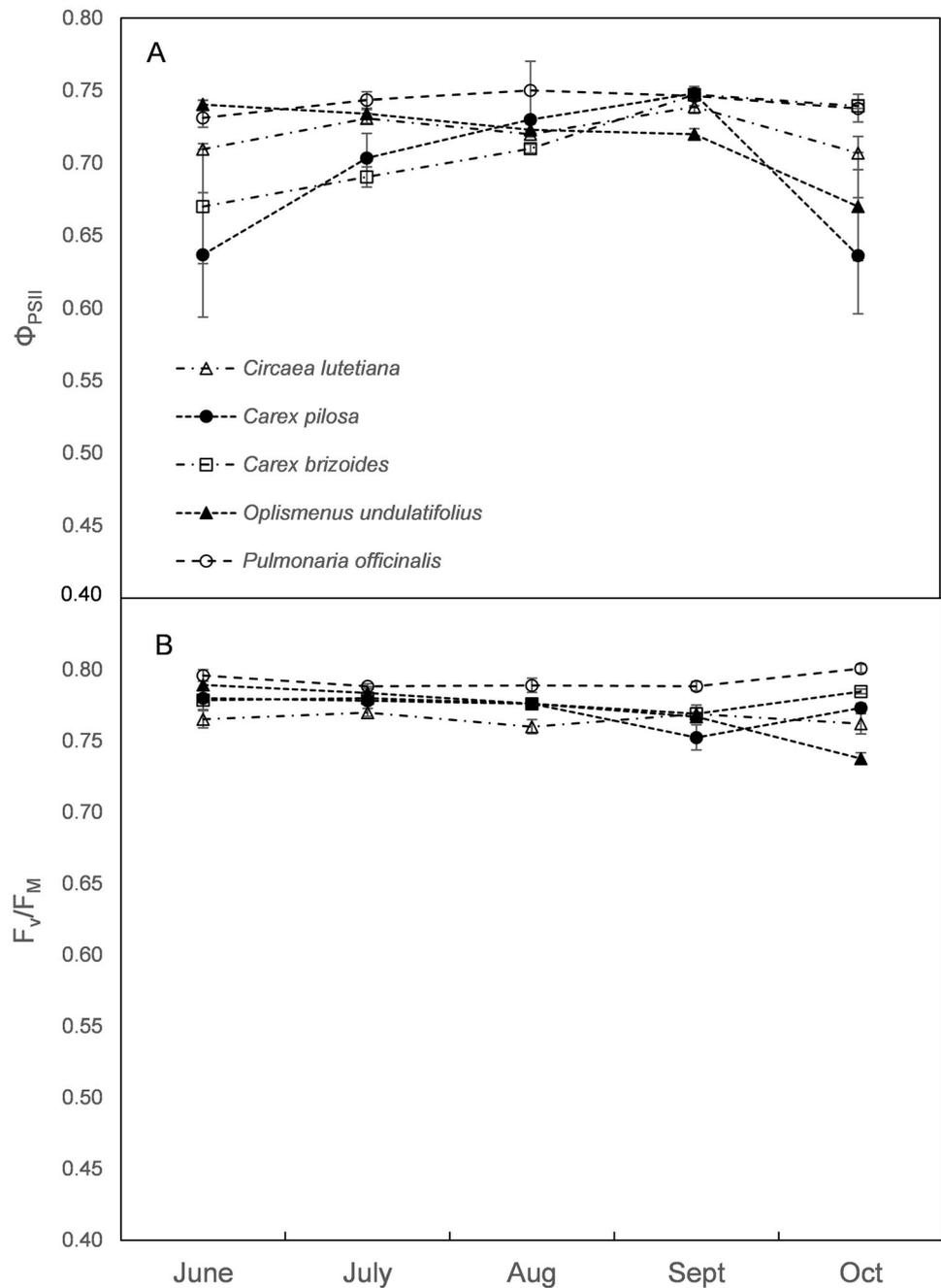
increase observed in October in *C. pilosa*, *P. officinalis* and *C. brizoides* when approaching overstorey canopy senescence, while the annual species (*C. lutetiana* and *O. undulatifolius*) show a more steady  $P_N$  trend during the study period, without the further increase in autumn. Therefore, the analyzed understory herbaceous species differ in their photosynthetic characteristics in way that are consistent with differences in their above-ground phenologies according to the results of Rothstein and Zak (2001).

Concerning the first question analyzed in this study and related to the  $g_m$  response to environmental conditions, we did not find any significant relationship between  $g_m$  and the considered environmental variables (i.e. atmospheric  $\text{CO}_2$  concentration,  $T_{\text{air}}$  and PPFD). In our study, the absence of these correlations can be related to the fact that the environmental changes occurring in the forest understory are not large enough to determine  $g_m$  variations according to the results of Nascimento and Marenco (2013). This is because the forest understory experiences damped environmental fluctuations, due to shelter by the overstorey tree canopy (Fotelli et al. 2003), that, in this case is characterized by a tree layer constituted by *Quercus robur* L., *Robinia pseudoacacia* L., *Populus nigra* L. and *Populus alba* L., and by an underlying layers with the presence of the species of the dominant tree layer and by *Corylus avellana* L., *Acer campestre* L., *Prunus padus* L. and *Crataegus monogyna* Jacq. (Granata et al. 2019). Moreover, it is worth noting that, the  $g_s$  values in

the forest tree layer (i.e. high-light level), during the more favorable environmental condition, ranges from  $0.263 \pm 0.017 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$  in *P. alba* and *P. nigra* (mean value), followed by *R. pseudoacacia* [ $0.221 \pm 0.005 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$ ] and by *Q. robur* [ $0.196 \pm 0.009 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$ ]. In the underlying layers (i.e. lower light level), the  $g_s$  value varies from  $0.195 \pm 0.006 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$  in *R. pseudoacacia*, followed by *A. campestre* [ $0.170 \pm 0.005 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$ ], *C. avellana* [ $0.160 \pm 0.003 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$ ], *C. monogyna* [ $0.154 \pm 0.006 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$ ], *P. padus* [ $0.117 \pm 0.006 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$ ], and by *Q. robur*, *P. alba* and *P. nigra* [ $0.101 \pm 0.006 \text{ mol}(\text{H}_2\text{O}) \text{m}^{-2} \text{s}^{-1}$ , mean value] (data not published). These results allow us to highlight a decrease in  $g_s$  rates between the same species present in different light conditions (tree layer vs underlying tree layer).

To understand the similarity between the species, based on the analyzed morphological and physiological variables, we carried out a Principal Component Analysis, which identifies three groups for the five considered species: one group is characterized by *C. lutetiana* and *O. undulatifolius*, the other by *C. pilosa* and *P. officinalis*, while *C. brizoides* is positioning far from the others species. In particular, the first component (i.e. PC1) which explains the major variation among the species is significantly correlate to the leaf morphological traits (i.e. LMA and LTD). Thus, we can conclude that the similarity and then the association between these species is mainly drive

**Fig. 3** Trend of **a** the actual PSII photochemical efficiency ( $\Phi_{PSII}$ ) and **b** the maximal quantum efficiency of PSII ( $F_v/F_m$ ) during the study period in the five considered species. Mean values ( $\pm$  SE) are shown ( $n = 10$ )

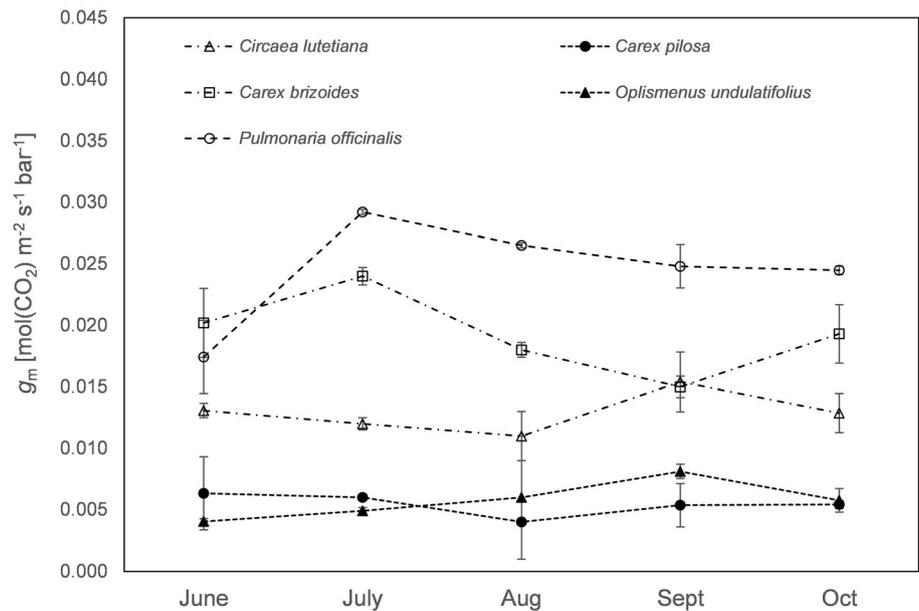


by the morphological leaf traits. In particular, among the considered species, *C. brizoides* is characterized by the highest values of both LMA and LTD ( $5.85 \pm 0.48 \text{ mg cm}^{-2}$  and  $1024.1 \pm 65.1 \text{ mg cm}^{-3}$ , respectively), while *C. lutetiana* and *O. undulatifolius* have the lowest LMA ( $1.78 \pm 0.32 \text{ mg cm}^{-2}$ , mean value). The other two species have a 31% lower LMA compared to *C. brizoides*, but due to a highest LT, they have a 65% lower LTD ( $406.3 \pm 16.0 \text{ mg cm}^{-3}$  in *C. pilosa* and  $319.7 \pm 13.8 \text{ mg cm}^{-3}$  in *P. officinalis*).

Furthermore, the significant relationship between the PC1 and  $g_m$  highlights that these morphological leaf traits are good predictors for the  $g_m$  variations across the considered species. In fact, leaf structural features are believed to play a central role in determining  $g_m$  (Hassiotou et al. 2009).

Thus, if we can suppose that these associations are driven by leaf structural traits and we found an association between one dicot and one monocot (i.e. *C. lutetiana* and *O. undulatifolius* in one group and *C. pilosa* and *P. officinalis* in the other group) and in turn these

**Fig. 4** Trend of mesophyll conductance ( $g_m$ ) during the study period in the five considered species. Mean values ( $\pm$  SE) are shown ( $n = 5$ )

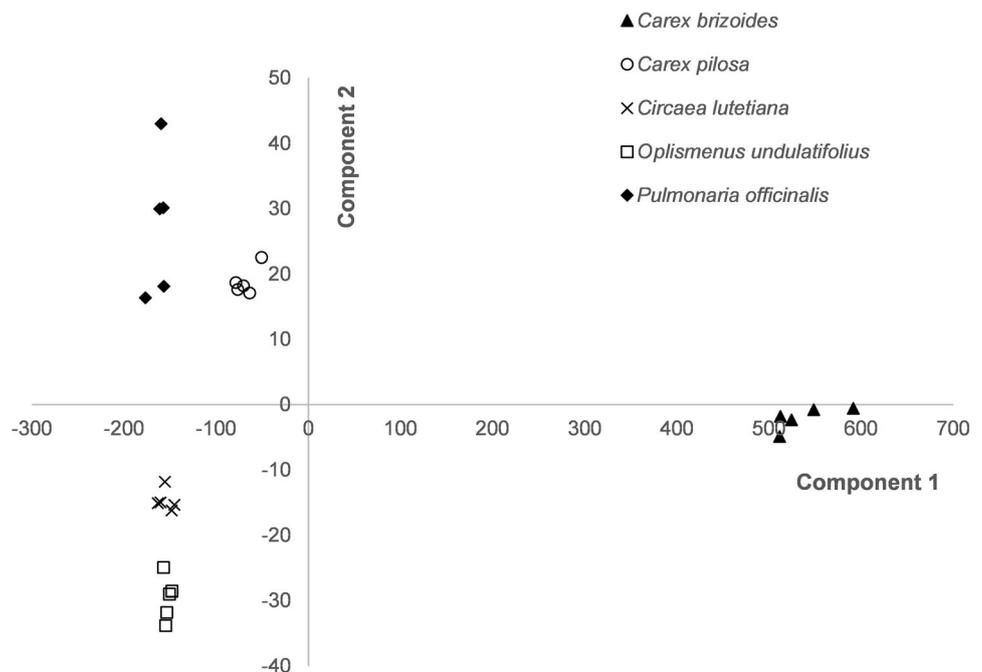


**Table 2** Regression analysis between mesophyll conductance ( $g_m$ ) and air temperature ( $T_{air}$ ), between  $g_m$  and photosynthetic photon flux density (PPFD) and between  $g_m$  and atmospheric  $CO_2$  concentration [ $CO_2$ ]. Regression analysis equation determination coefficient ( $R^2$ ) are shown

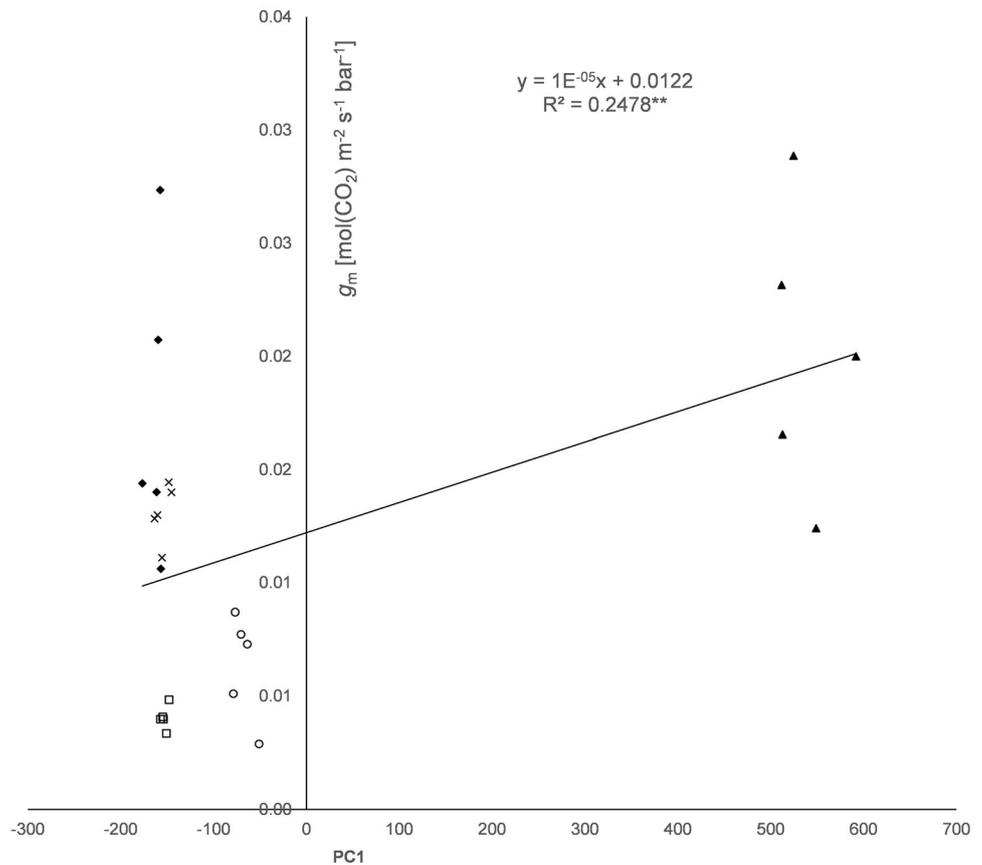
	Regression equation	$R^2$
$g_m$ versus $T_{air}$	$y = 0.004x + 0.0037$	0.00208
$g_m$ versus PPFD	$y = -1 e^{-0.5x} + 0.0141$	0.0148
$g_m$ versus [ $CO_2$ ]	$y = -9 e^{-0.6x} + 0.0171$	0.0011

morphological leaf traits are the best predictors of  $g_m$  variation in these species, we can conclude that the  $g_m$  variations are more linked to the functional group rather than reflecting an evolutionary trend. In fact, at the level of the functional groups, *C. lutetiana* and *O. undulatifolius* can be considered as annual species, while *C. pilosa* and *P. officinalis* are perennial species and *C. brizoides* seem to have an intermediate behavior between the two groups.

**Fig. 5** Principal component analysis (PCA) carried out using morphological (LMA, LTD and LT) and physiological ( $P_N$ ,  $R_D$ ,  $g_m$ ,  $g_s$ ,  $F_v/F_m$ ,  $\Phi_{PSII}$ , ETR) leaf traits for the five considered species. The 1st component, accounting for 99.4% of the total variance, was positively related to LMA and LTD. The 2nd component explained 0.62% of the total variance and it was correlated to LT



**Fig. 6** Linear relationship between the first principal component combining LMA and LTD (PC1) and the mesophyll conductance [ $g_m$ , mol(CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup> bar<sup>-1</sup>] across the five considered species. The equations as well as their R<sup>2</sup> are shown. (\*\* $p \leq 0.01$ )



**Acknowledgements** The work was supported by the ‘Natural Reserve Bosco Siro Negri’ funded by the Ministry of the Environmental Protection of Land and Sea of Italy.

**Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no conflict of interest.

**References**

Atkin OK, Evans JR, Ball MC, Siebke K (1998) Relationships between the inhibition of leaf respiration by light and enhancement of leaf dark respiration following light treatment. *Aust J Plant Physiol* 25:437–443

Bernacchi CJ, Portis AR, Nakano H et al (2002) Temperature response of mesophyll conductance. Implications for the determination of rubisco enzyme kinetics and for limitations to photosynthesis in vivo. *Plant Physiol* 130:1992–1998

Bernacchi CJ, Morgan PB, Ort DR, Long SP (2005) The growth of soybean under free air [CO<sub>2</sub>] enrichment (FACE) stimulates photosynthesis while decreasing in vivo Rubisco capacity. *Planta* 220:434–446

Bongi G, Loreto F (1989) Gas-exchange properties of salt stressed olive (*Olea europaea* L.) leaves. *Plant Physiol* 90:1408–1416

Cai ZQ, Slot M, Fan ZX (2005) Leaf development and photosynthetic properties of three tropical tree species with delayed greening. *Photosynthetica* 43:91–98

Cano F, Sánchez-Gómez D, Rodríguez-Calcerrada J, Warren CR, Gil L, Aranda I (2013) Effects of drought on mesophyll conductance and photosynthetic limitations at different tree canopy layers. *Plant Cell Environ* 36:1961–1980

Castagneri D, Garbarino M, Nola P (2013) Host preference and growth patterns of ivy (*Hedera helix* L.) in a temperate alluvial forest. *Plant Ecol* 214:1–9

Catoni R, Granata MU, Sartori F, Varone L, Gratani L (2015a) *Corylus avellana* responsiveness to light variations: morphological, anatomical and physiological leaf trait plasticity. *Photosynthetica* 53(1):35–46

Catoni R, Gratani L, Sartori F, Varone L, Granata MU (2015b) Carbon gain optimization in five broadleaf deciduous trees in response to light variation within the crown: correlations among morphological, anatomical and physiological leaf traits. *Acta Bot Croat* 74(1):71–94

De Lucia EH, Whitehead D, Clearwater MJ (2003) The relative limitation of photosynthesis by mesophyll conductance in co-occurring species in a temperate rainforest dominated by the conifer *Dacrydium cupressinum*. *Funct Plant Biol* 30:1197–1204

Diaz-Espejo A, Nicolás E, Fernández JE (2007) Seasonal evolution of diffusional limitations and photosynthetic capacity in olive under drought. *Plant Cell Environ* 30:922–933

Evans JR, Terashima I (1988) Photosynthetic characteristics of spinach leaves grown with different nitrogen treatments. *Plant Cell Physiol* 29:157–165

Flexas J, Ribas-Carbo M, Diaz-Espejo A, Galmes J, Medrano H (2008) Mesophyll conductance to CO<sub>2</sub>: current knowledge and future prospects. *Plant Cell Environ* 31:602–621

Flexas J, Barbour MM, Brendel O, Cabrera HM, Carriqui M, Diaz-Espejo A, Douthe Dreyer E, Ferrio JP, Gago J, Galle A, Galmes

- J, Kodama N, Medrano H, Niinemets U, Peguero-Pina JJ, Pou A, Ribas-Carbo M, Tomas M, Tosens T, Warren CR (2012) Mesophyll diffusion conductance to CO<sub>2</sub>: an unappreciated central player in photosynthesis. *Plant Sci* 193–194:70–84
- Fotelli MN, Rennenberg H, Holst T, Mayer H, Geßler A (2003) Carbon isotope composition of various tissues of beech (*Fagus sylvatica*) regeneration is indicative of recent environmental conditions within the forest understorey. *New Phytol* 159:229–244
- Genty B, Briantais JM, Baker NR (1989) The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochim Biophys Acta* 990:87–92
- Granata MU, Gratani L, Bracco F, Sartori F, Catoni R (2016) Carbon stock estimation in an unmanaged old-growth forest: a case study from a broad-leaf deciduous forest in the Northwest of Italy. *Int For Rev* 18(4):444–451
- Granata MU, Gratani L, Bracco F, Catoni R (2019) Carbon dioxide sequestration capability of an unmanaged old-growth broadleaf deciduous forest in a Strict Nature Reserve. *J Sustain Forest* 38:85–92
- Grassi G, Magnani F (2005) Stomatal, mesophyll conductance and biochemical limitations to photosynthesis as affected by drought and leaf ontogeny in ash and oak trees. *Plant Cell Environ* 28:834–849
- Hanba YT, Kogami H, Terashima I (2002) The effect of growth irradiance on leaf anatomy and photosynthesis in Acer species differing in light demand. *Plant Cell Environ* 25:1021–1030
- Harley PC, Loreto F, di Marco G, Sharkey TD (1992) Theoretical consideration when estimating the mesophyll conductance to CO<sub>2</sub> flux by analysis of the response of photosynthesis to CO<sub>2</sub>. *Plant Physiol* 98:1429–1436
- Hassiotou F, Ludwig M, Renton M, Veneklaas EJ, Evans JR (2009) Influence of leaf dry mass per area, CO<sub>2</sub>, and irradiance on mesophyll conductance in sclerophylls. *J Exp Bot* 60(8):2303–2314
- Jones HG (1973) Moderate-term water stresses and associated changes in some photosynthetic parameters in cotton. *New Phytol* 72:1095–1105
- Krall JP, Edwards GE (1992) Relationship between photosystem II activity and CO<sub>2</sub> fixation in leaves. *Physiol Plantarum* 86:180–187
- Kumar A, Turner NC, Singh DP et al (1999) Diurnal and seasonal patterns of water potential, photosynthesis, evapotranspiration and water use efficiency of clusterbean. *Photosynthetica* 37:601–607
- Laisk A, Eichelmann H, Oja V, Rasulov B, Padu E, Bichele I, Pettai H, Kull O (2005) Adjustment of leaf photosynthesis to shade in a natural canopy: rate parameters. *Plant Cell Environ* 28:375–388
- Massacci A, Nabiev SM, Pietrosanti L et al (2008) Response of the photosynthetic apparatus of cotton (*Gossypium hirsutum*) to the onset of drought stress under field conditions studied by gas-exchange analysis and chlorophyll fluorescence imaging. *Plant Physiol Biochem* 46:189–195
- Miyazawa SI, Terashima I (2001) Slow development of leaf photosynthesis in an evergreen broad-leaved tree, *Castanopsis sieboldii*: relationships between leaf anatomical characteristics and photosynthetic rate. *Plant Cell Environ* 24:279–291
- Motta R, Nola P, Berretti R (2009) The rise and fall of the black locust (*Robinia pseudoacacia* L.) in the “Siro Negri” Forest Reserve (Lombardy, Italy): lessons learned and future uncertainties. *Ann For Sci* 66:410–419
- Nascimento HCS, Marengo RA (2013) Mesophyll conductance variations in response to diurnal environmental factors in *Myrcia paivae* and *Minuartia guianensis* in Central Amazonia. *Photosynthetica* 51:457–464
- Park SY, Furukawa A (1999) Photosynthetic and stomatal responses of two tropical and two temperate trees to atmospheric humidity. *Photosynthetica* 36:181–186
- Piel C, Frak E, Le Roux X, Genty B (2002) Effect of local irradiance on CO<sub>2</sub> transfer conductance of mesophyll in walnut. *J Exp Bot* 53:2423–2430
- Rothstein DE, Zak DR (2001) Photosynthetic adaptation and acclimation to exploit seasonal periods of direct irradiance in three temperate, deciduous-forest herbs. *Funct Ecol* 15:722–731
- Taylor RJ, Pearcy RW (1976) Seasonal patterns of the CO<sub>2</sub> exchange characteristics of understorey plants from a deciduous forest. *Can J Bot* 54:1094–1103
- Tazoe Y, von Caemmerer S, Badger MR, Evans JR (2009) Light and CO<sub>2</sub> do not affect the mesophyll conductance to CO<sub>2</sub> diffusion in wheat leaves. *J Exp Bot* 60:2291–2301
- Tosens T, Niinemets Ü, Vislap V, Eichelmann H, Castro Díez P (2012) Developmental changes in mesophyll diffusion conductance and photosynthetic capacity under different light and water availabilities in *Populus tremula*: how structure constrains function. *Plant Cell Environ* 35:839–856
- von Caemmerer S, Evans JR (1991) Determination of the average partial pressure of CO<sub>2</sub> in chloroplasts from leaves of several C3 plants. *Aust J Plant Physiol* 18:287–305
- Warren CR, Ethier GH, Livingston NJ, Grant NJ, Turpin DH, Harrison DL, Black TA (2003) Transfer conductance in second growth Douglas-fir (*Pseudotsuga menziesii* (Mirb.) Franco) canopies. *Plant Cell Environ* 26:1215–1227
- Warren CR, Löw M, Matyssek R, Tausz M (2007) Internal conductance to CO<sub>2</sub> transfer of adult *Fagus sylvatica*: variation between sun and shade leaves and due to free-air ozone fumigation. *Environ Exp Bot* 59:130–138
- Yamori W, Noguchi K, Hanba YT, Terashima I (2006) Effects of internal conductance on the temperature dependence of the photosynthetic rate in spinach leaves from contrasting growth temperatures. *Plant Cell Physiol* 47:1069–1080

**Publisher’s Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.